Amendments to the specification:

Rewrite page 36, line 38, through page 38, line 9:

First, a standard curve was generated to determine the efficiency of the PCR with specific primers for the gene coding for SCN2A:
5'- TGCAGCAAACAAGGAAGAGCT -3' (SEQ ID NO: 4) and
5'-CGGGCTTTTCATCATTGAGTG 3' (SEQ ID NO: 5).
PCR amplification (95 °C and 1 sec, 56 °C and 5 sec, and 72 °C and 5 sec) was performed in a volume of 20 µl containing Lightcycler-FastStart DNA Master SYBR Green I mix (contains FastStart Taq DNA polymerase, reaction buffer, dNTP mix with dUTP instead of dTTP, SYBR Green I dye, and 1 mM $MgCl_2$, Roche), additional 3 mM MgCl2, 0,5 µM primers, and 2 µl of a cDNA dilution series (final concentration of 40, 20, 10, 5, 1 and 0.5 ng human total brain cDNA, Clontech). Melting curve analysis revealed a single peak at approximately 78.7°C with no visible primer dimers. Quality and size of the PCR product were determined with the DNA LabChip system (Agilent 2100 Bioanalyzer, Agilent Technologies). A single peak at the expected size of 74 bp for the SCN2A gene was observed in the electropherogram of the sample.

In an analogous manner, the PCR protocol was applied to determine the PCR efficiency of a set of reference genes which were selected as a reference standard for quantification. In the present invention, the mean value of five such reference genes was determined: (1) cyclophilin B, using the specific primers 5'-ACTGAAGCACTACGGGCCTG-3' (SEQ ID NO: 6) and 5'-AGCCGTTGGTGTCTTTGCC-3' (SEQ ID NO: 7) except for $MgCl_2$ (an additional 1 mM was added instead of 3 mM). Melting curve analysis revealed a single peak at approximately 87 °C with no visible primer dimers. Agarose gel analysis of the PCR product showed one single band of the expected size (62 bp). (2) Ribosomal protein S9 (RPS9), using the specific primers 5'-GGTCAAATTTACCCTGGCCA-3' (SEQ ID NO: 8) and 5'- TCTCATCAAGCGTCAGCAGTTC-3'

(SEQ ID NO: 9) (exception: additional 1 mM MgCl2 was added instead of 3 mM). Melting curve analysis revealed a single peak at approximately 85°C with no visible primer dimers. Agarose gel analysis of the PCR product showed one single band with the expected size (62 bp). (3) beta-actin, using the specific primers 5'- TGGAACGGTGAAGGTGACA-3' (SEQ ID NO: 10) and 5'-GGCAAGGGACTTCCTGTAA-3' (SEQ ID NO: 11). Melting curve analysis revealed a single peak at approximately 87°C with no visible primer dimers. Agarose gel analysis of the PCR product showed one single band with the expected size (142 bp). (4) GAPDH, using the specific primers 5'-CGTCATGGGTGTGAACCATG-3' (SEQ ID NO: 12) and 5'-GCTAAGCAGTTGGTGGTGCAG-3' (SEQ ID NO: 13). Melting curve analysis revealed a single peak at approximately 83°C with no visible primer dimers. Agarose gel analysis of the PCR product showed one single band with the expected size (81 bp). (5) Transferrin receptor TRR, using the specific primers 5'-GTCGCTGGTCAGTTCGTGATT-3' (SEQ ID NO: 14) and 5'-AGCAGTTGGCTGTTGTACCTCTC-3' (SEQ ID NO: 15). Melting curve analysis revealed a single peak at approximately 83°C with no visible primer dimers. Agarose gel analysis of the PCR product showed one single band with the expected size (80 bp).